Binding of ¹²⁵I-HCG to rat ovary homogenates at various stages of development

Age	Number of animals	s pg HCG bound/ Total	5 mg wet weight* Non-specific	Number of binding sites**		$ m K_D~(imes~10^{10})$
				moles/mg wet weight ($\times 10^{15}$)	moles/ovary ($\times~10^{15}$)	
Prenatal	40	40.8	35.4	0	0	
Newborn	250	45.0	40.6	0	0	-
5-day-old	250	40.7	33.4	0	0	
10-day-old	100	113.0	37.0	2.36	1.65	4.37
21-day-old	12	94.5	27.0	2.59	15.18	6.18
31-day-old	10	115.0	38.5	5.55	61.05	11.66
Spleen adult		41,3	36.5			

^{*}Determined with 4 ng 125I-HCG in the incubation assay. **Determined by Scatchard 22 analysis.

HCG was determined from samples containing a 1000fold excess of unlabelled hormone. The specific uptake of HCG was calculated from the difference between the radioactivity bound by samples incubated with 125I-HCG alone and the radioactivity bound nonspecifically. Iodination of purified HCG (biological activity of 11,000 IU/mg) was performed according to the modified method C of Leidenberger and Reichert 15 with a time of exposure to chloramine-T of 20 sec. Specific radioactivity of the ¹²⁵I-HCG was 30–50 μ Ci/ μ g.

Results and discussion. As can be seen from the table, ovarian tissues of prenatal, newborn and 5-day-old rats do not specifically bind radioactively labelled HCG. Specific binding of HCG is first observed in ovaries of 10day-old animals. These results are supported by the observation of Presl and Figarová⁸ that accumulation of injected radioactively labelled HCG is only demonstrable in ovaries of rats older than 7-8 days. The number of binding sites, as well as the dissociation constants (KD) of the receptor-hormone complex, are similar in 10-21day-old rats. Twice the number of binding sites as in the 21-day-old rat were demonstrable in rats 31 days of age. The values for K_D and number of binding sites in the 31day-old rat are largely consistent with those obtained by Lee and Ryan 14 in 35-day-old rats. It might be assumed that the lack of HCG binding in ovarian tissue of rats younger than 10 days is due to masking of the receptors by endogenous LH. This assumption can be abandoned, since specific HCG receptors are first demonstrable in ovarian tissue when the endogenous LH level has attained its maximum 3, 4, 16, 17.

These results strongly suggest that the HCG insensitivity of the early postnatal rat ovary is due to the lack of specific receptors. With the first appearance of the LH receptors in the ovary, the animals become sensitive to LH. Thus, in the female rat, in contrast to the male rat 18, LH receptor is not a constitutive protein. This difference between male and female is well understandable from a biological point of view. While steroidogenic activity of the testis is an absolute prerequisite for the development of the male phenotype, feminine differentiation occurs in the absence of gonadal steroids 19, 20. Only during the postnatal development do ovarian steroids become necessary. From the observation that in 10day-old female rats only the interstitial tissue of the ovary is steroidogenic 21, one might assume that interstitial cells are the first cells in the ovary to become endowed with LH receptors. The higher number of binding sites in the ovaries of 31-day-old rats compared with ovaries of 10day-old rats might indicate that besides interstitial cells also granulosa and theca cells are equipped with LH receptors at this developmental time.

- 15 F. Leidenberger and L. E. Reichert, Jr, Endocrinology 91, 135 (1972).
- N. B. Schwartz, C. H. Anderson, L. G. Nequin and C. A. Ely, in: Control of the Onset of Puberty, p. 367. Ed. M. M. Grumbach, G. D. Grave and F. E. Mayer. John Wiley and Sons, New York
- S. M. McCann, S. Ojeda and A. Negro-Vilar, in: Control of the Onset of Puberty, p. 1. Ed. M. M. Grumbach, G. D. Grave and F. E. Mayer. John Wiley and Sons, New York 1974.
- J. Frowein and W. Engel, Nature 249, 377 (1974).
- A. Jost, Phil. Trans. Roy. Soc., Lond. 259, 119 (1970).S. Ohno, U. Tettenborn and R. Dofuku, Hereditas 69, 107 (1971).
- W. D. Odell and R. S. Swerdloff, in: Control of the Onset of Puberty, p. 313. Ed. M. M. Grumbach, G. D. Grave and F. E. Mayer. John Wiley and Sons, New York 1974.
- 22 G. Scatchard, Ann. N. Y. Acad. Sci. 51, 660 (1949).

The effect of epinephrine and the hyperglycemic factor of the scorpion's cephalothoracic ganglionic mass (CTGM) on the phosphorylase activity of hepatopancreas of the scorpion, Heterometrus fulvipes C. Koch

K. Raghavaiah¹, M. Sreeramachandramurthy, R. Ramamurthi, P. Satyam and V. Chandrasekharam Department of Zoology, S. V. University and S.G. S. Arts College, Tirupati 517 501 (India), 8 September 1976

Summary. Injection of epinephrine and CTGM extract showed different effects on hepatopancreatic phosphorylase activity and levels of total carbohydrate and glycogen in Heterometrus fulvipes. The former hyperglycemic principle involves phosphorolysis of glycogen whereas the latter does not.

The role of neuroendocrine system in the metabolite regulation has been explored in Arachnida only to a limited extent². A hyperglycemic principle has been identified in the cephalothoracic ganglionic mass of the South Indian scorpion Heterometrus fulvipes 3, 4. The ver-

tebrate hormone, epinephrine also has been found to cause hyperglycemia in Heterometrus fulvipes.

The present communication examines the effect of epinephrine and scorpion's CTGM extract on the levels of total carbohydrates, glycogen and phosphorylase activity

Effect of injection of epinephrine or CTGM extract on the levels of total carbohydrate, glycogen, active (a) and total (ab) phosphorylase activities in the hepatopancreas of the scorpion H. fulvipes

Treatment	Total car	bohydrate	Glycogen		Phosphor a	rylases	ab		a • 100/al
1 Control 2 Epinephrine 3 CTGM	$6.95 \pm 1.12 \ (10) \ 5.85 \pm 1.21 \ (10) \ 9.69 + 1.29 \ (10)$		$3.31 \pm 0.88 (10)$ $2.82 \pm 0.80 (10)$ $4.34 + 0.76 (10)$		$8.0 \pm 1.25 (10)$ $22.2 \pm 3.22 (10)$ $8.5 + 0.95 (10)$		$28.2 \pm 3.26 (10)$ $46.2 \pm 6.82 (10)$		28.37 48.06
Comparison of mea	<u></u>				р	Percent	29.2 ± 4.	Percent	29.11
1–2	< 0.05	- 15.8	NS	- 14.8	< 0.001	+177.5	< 0.001	+63.8	
1-3 2-3	<0.001 - 0.001	+39.4 +65.6	<0.02 <0.001	$+31.1 \\ +53.9$	NS <0.001	+ 6.3 - 61.7	NS <0.001	+ 3.5 - 36.8	

Values for total carbohydrate and glycogen: mg/g dry weight; for phosphorylase: umole Pi/mg protein h. Number of determinations in brackets. NS, not significant.

of the hepatopancreas of the scorpion H. fulvipes. Collection, maintenance and details of preparation of experimental animals have been described earlier 4. Cephalothoracic ganglionic mass extracts in aqueous medium were injected into a batch of animals at the rate of one CTGM-equivalent (in 0.25 ml) per animal. For a second batch, epinephrine as adrenalin chloride (Parke-Davis, India), 1:1000, in saline solution was injected in 50 µgdose per animal. Controls received 0.25 ml of distilled water or saline. In the hepatopancreas of control, epinephrine injected and CTGM-extract injected scorpions, levels of total carbohydrates and glycogen were estimated by the anthrone method 5, and the phosphorylase activity was estimated by the method of Cori et al.6, and liberated inorganic phosphate was estimated according to Fiske and Subba Rau7. Protein content was estimated with Folin-Phenol reagent. 8.

The data and statistical treatment⁹ (table) reveals a fundamental difference between the mode of action of the vertebrate hormone and the scorpion's own hyperglycemic principle on the hepatopancreatic carbohydrate, glycogen and phosphorylase activity. Epinephrine leads to a significant (p < 0.05) decrease (-15.8%) of the total carbohydrate level of hepatopancreas, whereas CTGM extract injection results in a significant (p < 0.001) increase (+39.4%). The difference (+65.6%) between the total carbohydrate content of epinephrine- and CTGM extractinjected hepatopancreas also is significant (p < 0.001). Glycogen content shows a non-significant decrease on epinephrine treatment and CTGM extract leads to a significant (p < 0.001) elevation (+ 31.1%). The difference between the 2 treatments (+53.9%) is significant (p < 0.001), with regard to glycogen content also, as is the case with total carbohydrates. A 'prima facie' difference appears to exist in the modus operandi of hepatopancreatic carbohydrate metabolization between the 2 hormonal principles under consideration. This is further illustrated by the different influences exerted by these principles on the hepatopancreatic phosphorylase activity. Epinephrine elevates the levels of both active (a) and total (ab) phosphorylase levels significantly. Also this hormone brings about an alteration in the ratio of active to total phosphorylase. This ratio is elevated by 69.4% over control. But CTGM extract has no appreciable action on this enzyme system. Yet both these principles lead to development of significant levels of hyperglycemia4.

The principle of CTGM-extract is not only different from epinephrine, as has been demonstrated here, but also from the hyperglycemic principle of the insect corpora cardiaca extract 10-13 which seems to influence the glycogenolytic pathway for the augmentation of haemolymph carbohydrate level through stimulation of fat body phosphorylase system. It is of interest to note that the millipede Spirostreptus asthenes possesses in its brain a hyperglycemic principle the action of which appears to be essentially similar to the CTGM-principle of the scorpion in its mode of action on the fat body carbohydrate levels and phosphorylase system 14.

In view of its significant accumulation under the influence of CTGM principle, the carbohydrate content of hepatopancreas of the scorpion may not be implicated as the important causal source for the hyperglycemia resulting under the same aegis. Under brain-extract injection in the millipede, Spirostreptus asthenes 14, and under CTGMextract injection in the scorpion, H. fulvipes⁴, decreases in total lipid content of fat body and hepatopancreas were 49.4% and 19.2% respectively.

- The authors are grateful to Proff. K.S.Swami and S. Nagaiah. An award of Junior Research Fellowship by the Council of Scientific and Industrial Research, New Delhi is acknowledged
- F. Sehnal, in: Chemical Zoology, vol. VI, chapt. 9, p. 307. Ed. M. Florkin and B. T. Scheer. Academic Press, New York and London 1971.
- K. Raghavaiah and R. Ramamurthi, All India Symp. comp. Physiol. 1975, p. 46.
- K. Raghavaiah and R. Ramamurthi, Ind. J. exp. Biol. (1975).
- N. V. Carroll, R. W. Longley and J. H. Roe, J. biol. Chem. 22, 583 (1956).
- G. T. Cori, B. Illingworth and P. J. Keller, in: Methods in Enzymology, vol. 1, p. 200. Ed. S. P. Colowick and N. O. Kaplan. Academic Press, New York 1955.
- C. H. Fiske and Y. Subbarau, J. biol. Chem. 66, 375 (1925).
- O. H. Lowry, R. J. Rosebrough, A. L. Farr and R. J. Randall, J. biol. Chem. 193, 265 (1951).
- S.K. Pillai and H.C. Sinha, in: Statistical methods for Biologists. Ramprasad & Sons, Agra, India, 1968.
- J. E. Steele, Gen. comp. Endocrinol. 3, 46 (1963).
- 11 W. S. Bowers and S. Friedman, Nature, Lond. 198, 685 (1963).
- A. W. Wiens and L. I. Gilbert, J. Insect Physiol. 13, 779 (1967). G. J. Goldsworthy, J. Insect Physiol. 15, 2131 (1969).
- P. Satyam, Ph. D. Thesis submitted to S. V. University, Tirupati 1976.